

Algorithm for Extracting Weak Bands Kinetics from the Transient Absorption Spectra of the *Rhodobacter sphaeroides* Reaction Center

PRIMARY PROCESSES OF CHARGE SEPARATION IN REACTION CENTERS

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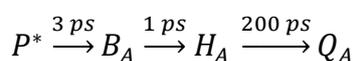
Abbreviations: ΔA , absorbance changes; ΔGauss , the fraction of Gauss absorption band changes; B_A , monomeric BChl in the active chain of cofactors; BChl, bacteriochlorophyll; H_A , bacteriopheophytin serving as an electron acceptor; P, primary electron donor; Q_A , primary quinone acceptor; Q_B , secondary quinone acceptor; RC, reaction center.

Abstract—An algorithm to extract kinetics of the ion radical bands from the strong absorption background in the transient absorption spectra of the *Rhodobacter sphaeroides* reaction centers upon femtosecond excitation of the primary electron donor is suggested. The rising kinetics of the transient absorption band at 1020 nm and the bleaching kinetics of the 545-nm band constructed using the proposed method are adequately fitted by the kinetic equations for sequential electron transfer from the excited primary donor to the B_A (monomeric bacteriochlorophyll) molecule, and then to the H_A (bacteriopheophytin serving as an electron acceptor) molecule with the rate constants of 3.5 ± 0.2 and 0.8 ± 0.1 ps, respectively. The kinetics of the bacteriochlorophyll absorption band at 600 nm shows both the ultrafast bleaching of the P_{870} dimer and slower bleaching of the B_A monomer due to its transition to the anion radical. The plotted kinetics of the ion radical bands is in agreement with the concentration profiles of the charge-separated states produced by the global target analysis of experimental data using the model of sequential electron transfer in the reaction centers.

Keywords: femtosecond spectroscopy, bacterial reaction centers, electron transfer

Photosynthetic reaction centers (RCs) of *Rhodobacter sphaeroides* are transmembrane protein complexes consisting of three subunits (L, M, and H) and ten electron transfer cofactors that mediate the ultrafast photophysical processes of the light energy transformation into the energy of charge-separated states. The electron transfer cofactors include two bacteriochlorophylls (BChls) forming the primary electron donor dimer (P), two monomeric bacteriochlorophylls (B_A and B_B), two bacteriopheophytins (H_A and H_B), two quinones (Q_A and Q_B), non-heme iron atom, and carotene molecule. The cofactors are organized into two pseudosymmetric chains A and B with only A chain being active [1, 2].

It is known that the transmembrane electron transfer from the excited donor P^* to the quinone molecule Q_A occurs in the RCs upon illumination. Nowadays it is generally accepted nowadays that the transmembrane electron transfer proceeds via a series of ultrafast reactions involving the B_A and H_A molecules from the active cofactor chain as intermediate electron acceptors. It is quite challenging to identify the $P^+B_A^-$ state experimentally because the rate of electron transfer to the B_A molecule is slower than the rate of electron transfer from B_A^- to H_A [3, 4]. This results in a low population of the $P^+B_A^-$ state, which does not exceed several tens of percent. Another problem relates to the fact that the spectrum of the $P^+B_A^-$ state is masked by the spectra of other states, including P^* . Identification of the minor band of the B_A^- anion radical at 1020 nm in the transient absorption spectra provided the basis for verification of the four-component model in which the B_A molecule is the primary electron acceptor [5, 6]. According to this model, the transmembrane electron transfer occurs from the excited donor P^* to the B_A molecule with the rate constant of ~ 3 ps, which is followed by its transfer to the H_A molecule with the rate constant of ~ 1 ps and next to the Q_A molecule with the rate constant of ~ 200 ps.



Recently, researchers investigating the primary processes of charge separation in the photosynthetic RCs often perform the global analysis of the transient absorption spectra measured with a femtosecond resolution [7] in order to describe an entire set of experimental data using mathematical and kinetic models.

This method allows approximating experimental data with the help of various models: (i) sequential electron transfer model [8-11]; (ii) model of electron transfer with

consideration of reversibility of the charge separation reactions [8-10]; and (iii) model of charge separation via two deactivation channels in the heterogeneous environment of protein cofactors in the RC [10]. It must be noted that extraction of the short-lived states using global analysis technique is difficult if the excitation pulse duration is sufficiently short and coherent effects are observed. Hence, in order to suggest an adequate physical model, global analysis requires additional information on the processes of charge separation.

In a number of cases, analysis of kinetics of absorption changes ΔA at a selected wavelength still remains relevant. However, such analysis can be complicated by the contribution of various overlapping spectral components. Exponential analysis of the complex kinetics of the absorption changes ΔA at 600 nm was suggested by Carter et al. [12] as one of some approaches for estimating the quantum yield of the $P^+H_B^-$ state formation in the mutant RC from *Rhodobacter capsulatus*. Another approach for solving the problem of overlapping bands is construction of kinetics of the B_A^- anion radical at 1020 nm using approximation of the spectral background by curved line over the wide wavelength range [13]. In this work, we suggest an alternative method for the extraction of kinetics of individual weak absorption bands, including the short-lived ones, followed by analysis of this kinetics within the framework of the sequential electron transfer model.

MATERIALS AND METHODS

Reaction centers (RCs) were isolated from *Rba. sphaeroides* as described previously [14]. After that, the RCs were suspended in 20 mM Tris-HCl (pH 8.0) containing 0.1% (v/v) Triton X-100 and concentrated to the optical density 0.5 at 870 nm (optical path length, 1 mm). The absorption spectra of the RCs were recorded with a UV1800 spectrophotometer (Shimadzu, Japan).

Detailed description of the femtosecond laser spectrometer has been presented by us in [15]. Femtosecond light pulses (~35 fs, ~3 mJ) were produced using a femtosecond MaiTai SP laser with a Spitfire Ace regenerative amplifier (Spectra-Physics, USA). The light with the wavelengths >850 nm was cut off from the continuum produced on the sapphire with an RG850 optical filter (Newport, USA) for excitation of the primary electron donor at 870 nm. The measuring beam was divided

into the probing and reference ones. The probing and reference pulses after the rotating cuvette (1 mm thickness) were first directed to a Spectra Pro 2300i spectrograph and next to a Pixis 400BR CCD camera (Princeton Instruments, USA) for recording the spectra in the 450-720 or 750-1100 nm ranges. The relative polarization of the excitation pulse was parallel to the measuring pulse. The cuvette rotation frequency and the excitation frequency (10 Hz) were selected in such a way that each excitation pulse illuminated a new portion of RCs. The intensity of the excitation pulses was attenuated to excite less than 20% RCs in the sample. All measurements were carried out at room temperature.

Global analysis of the transient absorption spectra was performed with the Glotaran program [16].

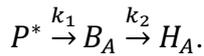
Algorithm for extraction of the spectral bands kinetics. The appearance of absorption bands in the transient spectra of the electron transport chain components at different delay times can be observed upon femtosecond excitation of the primary electron donor. Let us assume that the component absorption appears as a symmetric Gaussian band. The evolution of this spectral component in time is reflected in the kinetics of absorption changes ΔA in the absorption band maximum. On the other hand, one can always find **A** and **B** points in the vicinity of the absorption maximum that are located at equal distances δ from the maximum (Fig. 1, curve *I*). It is obvious that due to the symmetry of the band, the mean wavelength value for the points **A** and **B** is the wavelength of the absorption maximum, and the mean absorption for the points **A** and **B** is the midpoint of the **AB** segment. The difference in optical density at the absorption maximum and at the midpoint of the **AB** segment is the fraction of the Gaussian band amplitude with the kinetics of changes ΔA_{Gauss} being the same as the kinetics of absorption changes ΔA in the band maximum, but less in amplitude.

Various other spectral components contribute to the absorption within any absorption region, forming the background that changes in time. Hence, at any time delay, the optical density in the absorption band maximum constitutes the sum of the optical densities of the band and its background. That is why the time course of absorption changes ΔA at the absorption band maximum includes the kinetics of the band itself and of the background. Let us assume that the spectral background within a certain wavelength range is linear. In the case of arbitrary time delay and background represented by a straight line, the supplementary straight line that is parallel to the background will cross the Gaussian absorption bands in points **A'** and **B'** located at

equal distances from the absorption band maximum. It is easy to see that for different delay times, the segment $A'B'$ always remains parallel to the background, and in the case when the band is absent, it is located on the background (Fig. 1, curve 2). If the vector half-sum of the ΔA kinetics at the points A' and B' is subtracted from the ΔA kinetics at the absorption band maximum, we obtain as a result a vector difference that represents the same fraction of the spectral band for each time delay. In this case, the Δ Gauss kinetics constructed using the above described approach will be the kinetics of the band spectral fraction independent on the rate constant of the linear background amplitude and slope changes.

In the case when the spectral background displays a curvature within a certain wavelength range that does not change with time, this curvature will not affect the Δ Gauss kinetics but only shift it along the optical density axis. And only in the case when the background curving changes in time, it introduces distortions to the Δ Gauss kinetics, but these distortions are significantly smaller than the contribution of the entire spectral background as a whole.

Model of sequential electron transfer in RCs. According to the sequential model of charge separation in the RC complexes of purple bacteria, electron transfer from the excited primary donor P^* to the B_A molecule occurs with the rate constant k_1 under exposure to light within a picosecond time scale, and the next electron is transferred from B_A to bacteriopheophytin H_A with the rate constant k_2 :



At the initial time, $[P^*] = a$, $[B_A^-] = 0$, $[H_A^-] = 0$. Considering that a single electron is transferred during the elementary act of charge separation, the mass balance equation is: $[P^*] + [B_A^-] + [H_A^-] = a$.

The system of differential equations for the electron transfer components is the following:

$$\frac{d[P^*]}{dt} = -k_1[P^*],$$

$$\frac{d[B_A^-]}{dt} = k_1[P^*] - k_2[B_A^-],$$

$$\frac{d[H_A^-]}{dt} = k_2[B_A^-].$$

Solving this system of equations allows producing equations of the kinetic curves for the concentrations of P*, B_A⁻, and H_A⁻:

$$[P^*] = a \cdot \exp(-k_1 t), \quad (1)$$

$$[B_A^-] = \frac{k_1 \cdot a}{k_2 - k_1} (\exp(-k_1 t) - \exp(-k_2 t)), \quad (2)$$

$$[H_A^-] = a \cdot \left\{ 1 + \frac{k_1 \exp(-k_2 t) - k_2 \exp(-k_1 t)}{k_2 - k_1} \right\}. \quad (3)$$

It can be seen from the presented equations that the P* concentration decreases exponentially with time, the concentration of B_A⁻ first increases reaching the maximum and then decreases to zero, and the concentration of H_A⁻ increases reaching the value *a*.

RESULTS AND DISCUSSION

The kinetics of the absorbance changes (ΔA) and in the fraction of ΔA_{Gauss} amplitude in the absorption region of the B_A⁻ anion radical in *Rba. sphaeroides* RCs is presented in Fig. 2. The kinetics of absorption changes ΔA_{1020} (Fig. 2a) is difficult to assign to the intermediate electron acceptor, as it does not decay in time up to several nanoseconds (data not shown) [4], which is related to the fact that other spectral components contribute significantly to the absorption at 1020 nm forming the background [17]. Indeed, negative absorption changes are observed at delay times less than one picosecond that are due to the stimulated emission at 935 nm, and positive changes due to the absorption of the primary electron donor cation radical are observed at the delay times of tens of picoseconds.

Evolution of the absorption band at 1020 nm occurs in the presence of spectral background changing with time (Fig. 2a, inset). The presence of this background at delay times up to tens of femtoseconds is due to the positive absorption at 1100 nm and

negative absorption of the stimulated emission at 935 nm. The sum of the positive and negative Gaussian functions that form the background for the narrow wavelength range around 1020 nm can be presented in the first approximation as a straight line. The amplitudes of these spectral bands decrease at the delay times less than tens of picoseconds, which results in the changes in the background slope; absorption of the primary electron donor cation radical appears, which causes an increase in the background amplitude. This consideration describes the evolution of the absorption band of the B_A^- anion radical occurring over the linear background with the slope and amplitude changing significantly with time. The kinetics of changes in the fraction of the spectral absorption band (ΔGauss) at 1020 nm constructed for the wavelength regions $\delta = \pm 15$ nm (curve 1), $\delta = \pm 10$ nm (curve 2), and $\delta = \pm 5$ nm (curve 3) is presented in Fig. 2b. The decrease of δ results in the decrease of the signal-to-noise ratio. It was shown (Fig. 2b) that the time course of the constructed ΔGauss kinetics did not depend on the level of experimental noise unavoidable during measuring of the transient absorption spectra. The ΔGauss kinetics at all δ values is characteristic for the intermediate electron acceptor. However, the decrease in the signal-to-noise ratio results in the increase of the error in the determination of time constants. This kinetics can be adequately approximated with the Eq. (2) for the intermediate electron acceptor that accepts electron with the rate constant of 3.5 ± 0.2 ps and transfers electron with the rate constant of 0.8 ± 0.1 ps, which is in good agreement with the rate constants for the B_A molecule in the model of sequential electron transfer [8, 11].

The kinetics of absorption changes ΔA and the kinetics of the ΔGauss band change in the region of Q_X transition in BChl at 600 nm are compared in Fig. 3. An instant increase in the absorption can be seen in the time course of the absorption change ΔA_{600} following the femtosecond excitation of P_{870} due to the excited state absorption of the primary electron donor, which is likely higher in the amplitude than the bleaching absorption band at 600 nm (Fig. 3a). Further, in the ΔA kinetics an exponential decay in the picosecond time scale with a lifetime of ~ 3.5 ps is observed, which is due to the decrease in the concentration of the excited primary electron donor P_{870}^* . At the same time, it can be seen in the inset in Fig. 3a that the absorption band at 600 nm bleaches completely in the transient absorption spectra within the interval of ~ 0.1 ps. Hence, the kinetics of absorption changes ΔA in the region of the Q_X transition of BChl at 600 nm is predominantly due to the evolution of the excited primary electron donor.

The kinetics of changes in the Δ Gauss band absorption at 600 nm constructed for the wavelength range $\delta = \pm 10$ nm represents evolution of the spectral band at 600 nm without the primary electron donor contribution (Fig. 3b). The kinetics shows instant (~ 60 fs) bleaching of the Q_X absorption band of the P_{870} dimer related to the primary donor transition to the excited state. Further, within the picosecond time scale and up to 2.5 ns (Fig. 3b, inset), the amplitude of the Δ Gauss bleaching does not change because the excited primary electron donor passes electron to the active chain of cofactors with the quantum yield close to one and is transformed into the cation radical form P_{870}^+ .

The additional increase of bleaching observed in the Δ Gauss kinetics in the early delay times has attracted our attention (Fig. 3b, inset). Considering that the level of the 600-nm band in the nanosecond time scale is due to absorption of the two BChl molecules of the special pair, this increase is likely associated with the third BChl. If one assumes that the dipole strength of the Q_X transitions of the four BChl molecules in the RC are comparable, it is possible to estimate the bleaching of this third BChl molecule at a level of only 20%, in contrast to the first two molecules. The increase of bleaching reaches the maximum at the delay time of 1.6 ps and decreases afterwards within ~ 10 ps (Fig. 3b). The kinetics of this additional component replicates the kinetics of the spectral band at 1020 nm (Fig. 2b) and is likely due to the bleaching of the Q_X absorption band of the B_A molecule during its transformation into the anion radical form. Comparison of Figs. 3a and 3b allows concluding that the femtosecond bleaching of the absorption band of the primary electron donor and the nanosecond bleaching of the B_A molecule due to the electron transfer to it are masked in the kinetics of ΔA at 600 nm by changes in the absorption of the excited primary electron donor, while in the Δ Gauss kinetics, these changes are clearly visible.

The kinetics of absorption changes ΔA in the Q_X region of bacteriopheophytin transition at 545 nm is presented in Fig. 4a. The fast increase in the absorption is due to the absorption of the excited primary electron donor. The decrease in the absorption related both to the decrease in the concentration of excited P_{870}^* and bleaching of the bacteriopheophytin H_A absorption band occurs within the picosecond time scale (Fig. 4a, inset) [18].

No fast processes are observed in the kinetics of the Δ Gauss band changes at 545 nm (Fig. 4b). Bleaching of the 545-nm band is observed over the picosecond time scale, which is due to the fact that the H_A molecule accepts an electron and is

transformed into the anion radical form H_A^- . A slight bend is observed in the kinetics of the band bleaching at 545 nm at the early time delays, which differs it from the exponential kinetics. Indeed, such kinetics can be perfectly fitted by the Eq. (3) for the secondary electron acceptor with the rate constants of 3.4 ± 0.2 and 0.7 ± 0.1 ps. All these observations allow concluding that the H_A molecule exhibits the properties of the secondary rather than primary electron acceptor. The fact that slight relaxation related to the electron transfer to the following quinone acceptor Q_A can be observed at time delays >15 ps is worth mentioning.

The kinetics of Δ Gauss at 545 nm constructed for the time range up to 2.5 ns is presented in the inset in Fig. 4b. It can be seen in this time scale, that the H_A molecule also acts as an intermediate acceptor of in the electron transition from the excited donor P^* to the quinone acceptor Q_A . The kinetics presented in the Fig. 4b inset can be fitted adequately with the Eq. (2) with the rate constants 3.5 ± 0.4 and 210 ± 10 ps.

The results produced in this study allows constructing the time dependences of the concentrations of P^* , $P^+B_A^-$, and $P^+H_A^-$, i.e., obtaining experimentally derived kinetic curves of the states with separated charges (Fig. 5a). The kinetics of absorption changes ΔA at 930 nm (Fig. 5a, curve 1) due to the stimulated emission decay reflects evolution of the primary electron donor excited state. The kinetics of the Δ Gauss spectral band at 545 nm (Fig. 5a, curve 3) represents evolution of the $P^+H_A^-$ state. We have normalized these curves to one, because the quantum yield of the $P^+H_A^-$ charge separation in the RC is close to one. The kinetics of the Δ Gauss spectral band at 1020 nm (Fig. 5a, curve 2) represents evolution of the $P^+B_A^-$ state. This kinetics has been normalized to 0.2 as the Q_X absorption band of the B_A molecule at 600 nm bleaches only by $\sim 20\%$.

At the same time, we performed global analysis of the entire set of experimental data in the frameworks of several kinetic models: (i) model of electron transfer with sequentially decreasing rate constants $k_1 > k_2 > k_3$ [9]; (ii) model of electron transfer with rate constants of forward reactions $k_{12} < k_{23} > k_{34}$ considering the reaction of charge recombination in the $P^+H_A^-$ state occurring via the B_A molecule with recombination rate constants $k_{21} = 1/12$ ps and $k_{32} = 1/21$ ps [8, 10], and (iii) model of sequential electron transfer with the rate constants $k_1 < k_2 > k_3$ without consideration of reverse reactions [11]. The concentration profiles of the P^* , $P^+B_A^-$, and $P^+H_A^-$ states were derived for all these models. It was found that the profile of the $P^+B_A^-$ state concentration for the model with sequentially decreasing rate constants at the maximum reaches $\sim 60\%$ rather than 20%. For the model of electron transfer considering reverse reactions, the occupancy of

the $P^+B_A^-$ state is maintained for an extended time period, and this state can be observed within the time interval up to several hundred picoseconds. Good agreement of the experimentally measured kinetics of the weak absorption bands with the modeled concentration profiles of the charge separated states was observed only for the sequential model of electron transfer with the rate constants of 3, 1, and 200 ps without considering reverse reactions.

Comparison of Figs. 5a and 5b reveals that the kinetics of weak absorption bands ΔA_{Gauss} at 1020 and 545 nm and the ΔA kinetics at 930 nm constructed without any a priori models accurately show the concentration profiles of the charge separated states for the sequential four-component model of the electron transfer in the RC.

This allows concluding that the kinetic curves constructed using the algorithm suggested in this work are only slightly distorted by the contribution of the accompanying spectral components and satisfactorily reproduce the evolution of the concentrations of the charge separated states. These data further confirm the results obtained previously by our research group [6, 13] and in the recent study by Dominguez et al. [11] according to which sequential homogenous electron transfer occurs in the RC that involves the B_A molecule as the primary electron acceptor [19]. This is also in agreement with the fact that both the kinetics of the 1020-nm band and the profile of the $P^+B_A^-$ state concentration in this model decay within the time interval up to ~ 10 ps. Meanwhile, in the model of reverse electron transfer, the occupancy of the $P^+B_A^-$ state is maintained for up to several hundreds of picoseconds [8, 10]. The presented data also indicate that following excitation of the electron donor with a pulse duration ~ 30 fs, the charge separation in the RC at room temperature proceeds according to the nonadiabatic mechanism.

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Conflict of Interest

The authors declare no conflict of interest in financial or any other sphere.

Ethical Approval

This article does not contain any studies with human participants or animals performed by any of the authors.

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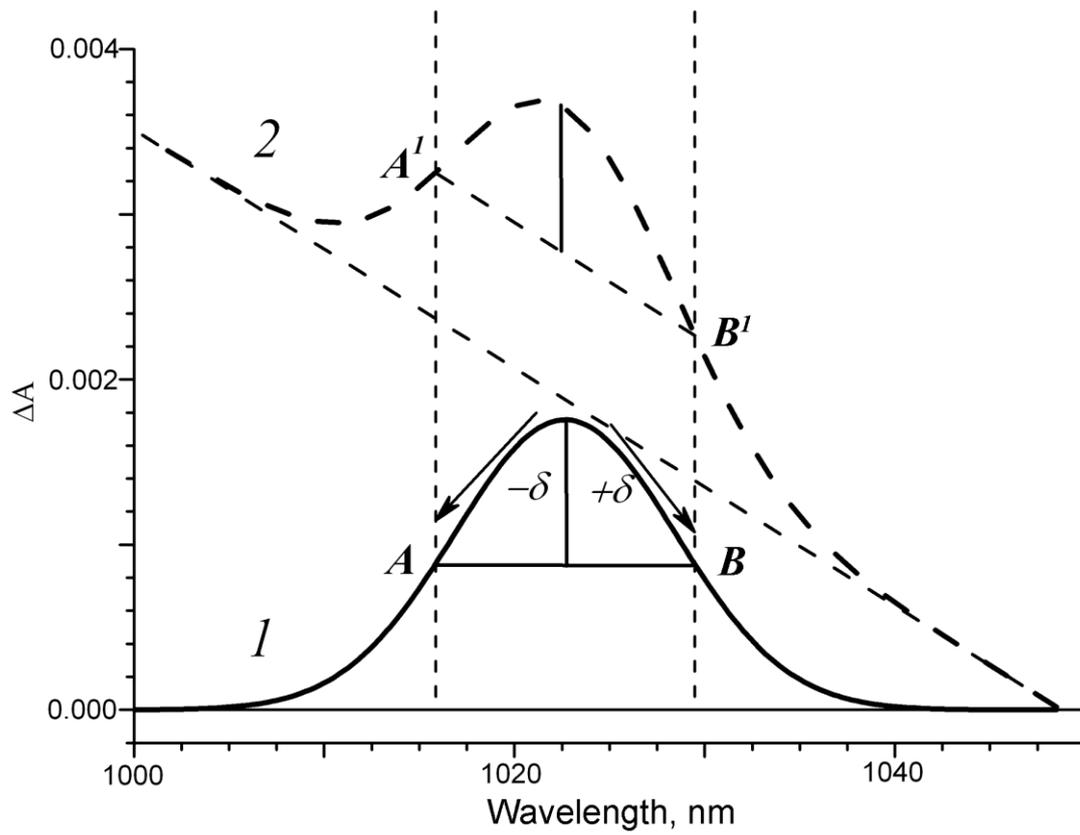


Fig. 1. Model of symmetric Gaussian band in the transient absorption spectra: 1) without background; 2) with linear spectral background.

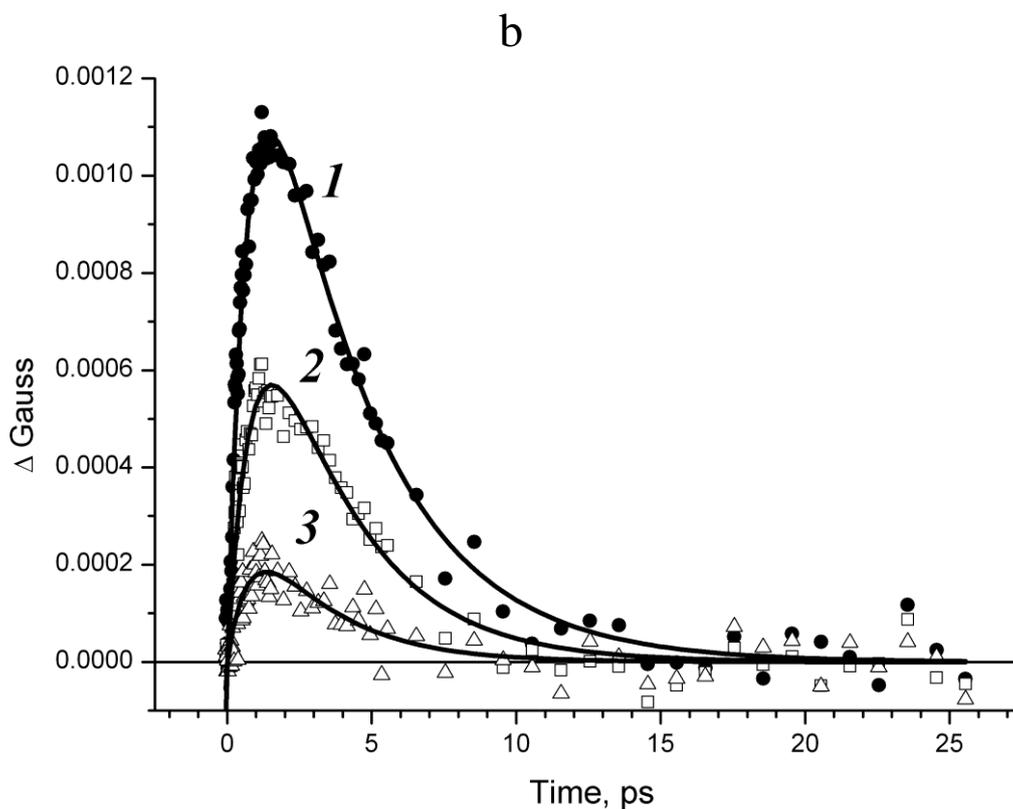
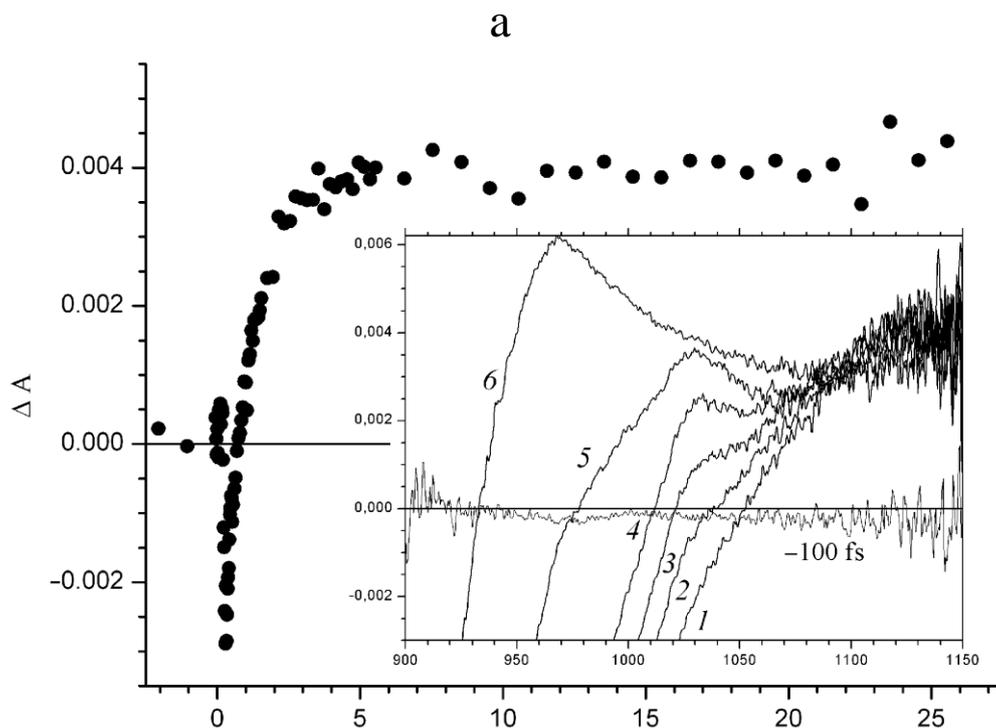


Fig. 2. a) Kinetics of absorption changes ΔA at 1020 nm. Inset, transient absorption spectra recorded at delay times 0.3 (1), 0.53 (2), 0.85 (3), 1.5 (4), 3.7 (5), and 10.6 ps (6). b) Kinetics of the fraction of Δ Gauss spectral band at 1020 nm constructed at the wavelength ranges $\delta = \pm 15$ nm (1), $\delta = \pm 10$ nm (2), and $\delta = \pm 5$ nm (3). Approximation of the kinetics with the Eq. (2) is presented by solid lines.

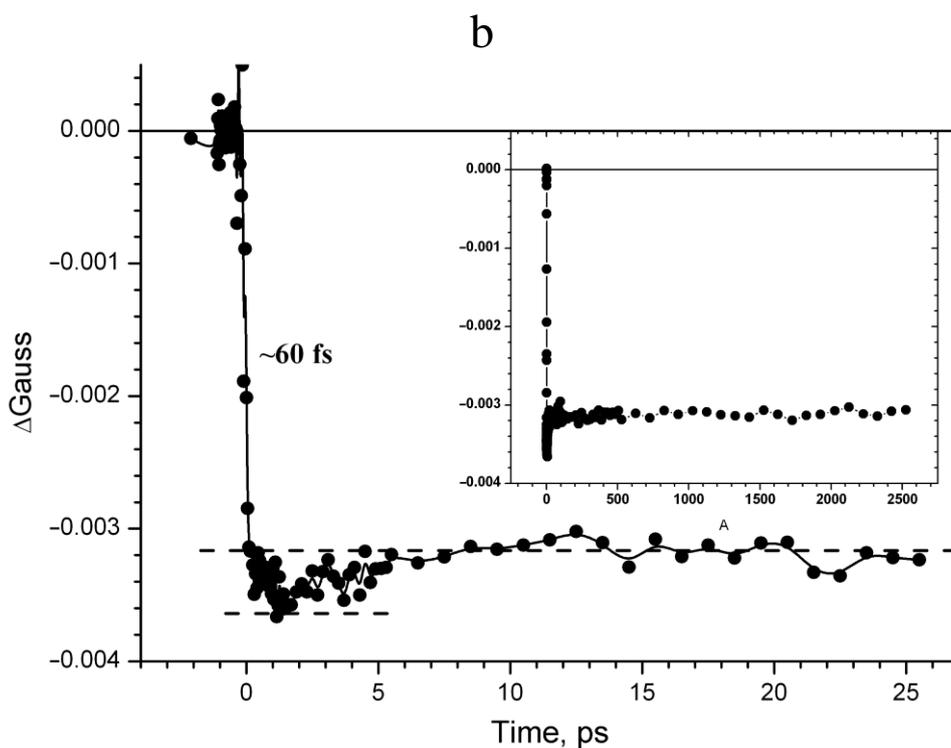
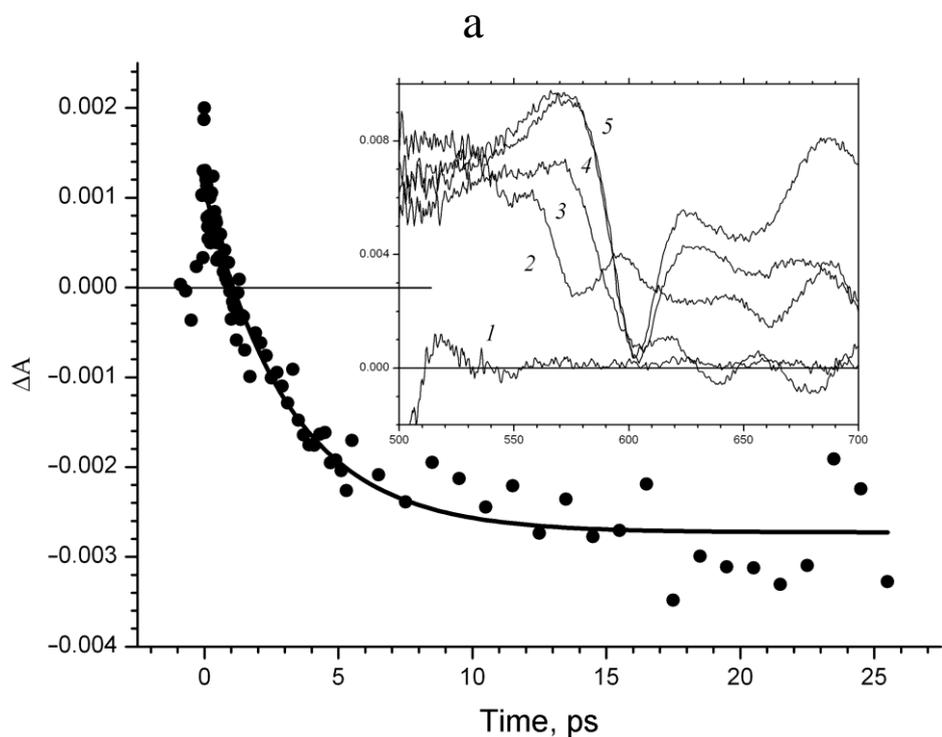


Fig. 3. a) ΔA kinetics at 600 nm recorded in time interval from -2 to 27.5 ps. Exponential approximation of the kinetics is depicted by solid line. Inset, transient absorption spectra recorded at delay times -0.5 (1), -0.15 (2), -0.05 (3), 0.05 (4), and 0.1 ps (5). b) Kinetics of changes in the fraction of the Δ Gauss spectral band at 600 nm (wavelength range $\delta = \pm 10$ nm) constructed for times from -2 to 27.5 ps. Inset, Δ Gauss kinetics at 600 nm constructed for delay times from -20 ps to 2.5 ns.

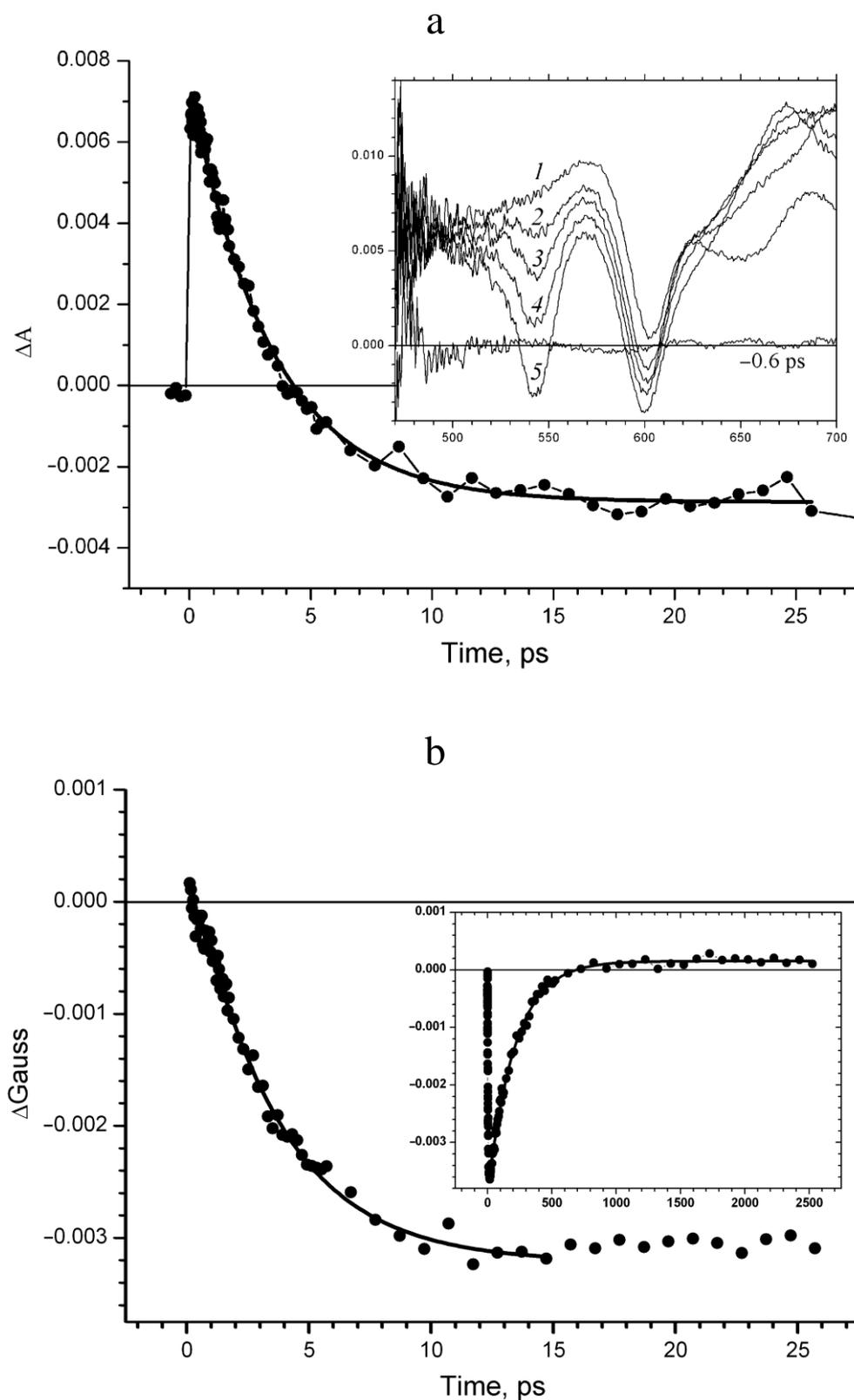


Fig. 4. a) ΔA kinetics at 545 nm recorded for times from -2 to 27.5 ps. Inset, transient absorption spectra recorded for the delay times 0.1 (1), 1.0 (2), 2.0 (3), 3.0 (4), and 8.0 ps (5). b) Kinetics of changes of the fraction of ΔG auss spectral band at 545 nm (wavelength range $\delta = \pm 10$ nm) constructed for times from -2 to 27.5 ps. Approximation of the kinetics with Eq. (3) is depicted with solid line. Inset, ΔG auss kinetics at 545 nm constructed for times from -20 ps to 2.5 ns.

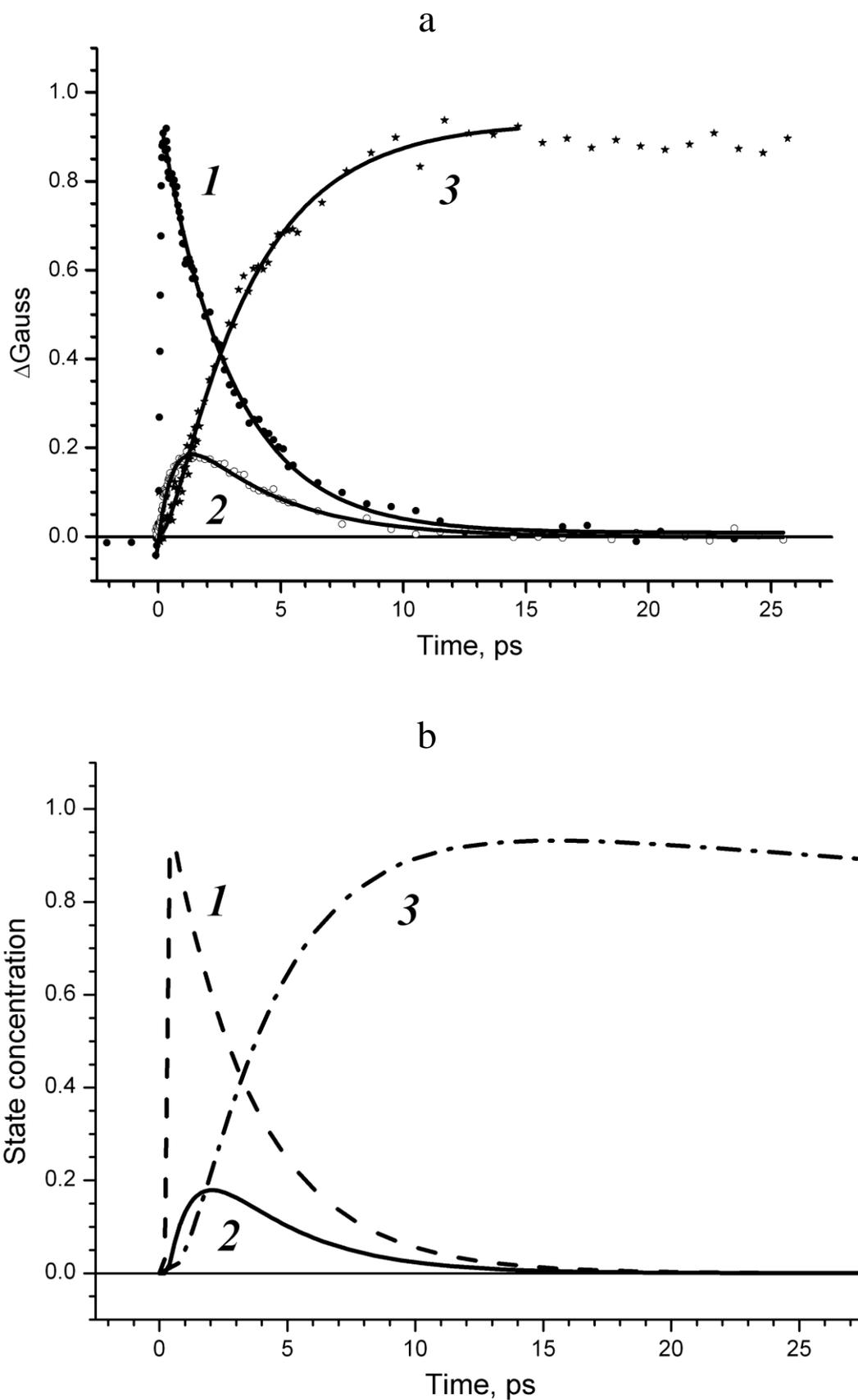


Fig. 5. a) Kinetics of ΔA at 928 nm (1) and ΔGauss at 1020 nm (2) and 545 nm (3) constructed for times from -2 to 27.5 ps. b) Concentration profiles of the P^* (1), $P^+B_A^-$ (2), and $P^+H_A^-$ (3) states obtained using global analysis of experimental data.